

ASPERTOXIN, A HYDROXY DERIVATIVE OF O-METHYLSTERIGMATOCYSTIN
FROM AFLATOXIN-PRODUCING CULTURES OF ASPERGILLUS FLAVUS

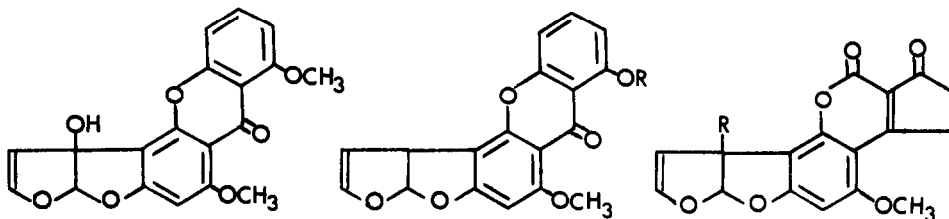
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A preliminary report from our laboratories¹ described the isolation and biological activity of a new toxic metabolite from an aflatoxin-producing culture of Aspergillus flavus, and the name "aspertoxin" was tentatively assigned to the compound. On the basis of further investigations of this substance, we propose that aspertoxin is a hydroxy derivative of another metabolite of A. flavus, O-methylsterigmatocystin (II, R=CH₃),² and is represented by formula I. Aspertoxin also bears a close resemblance to aflatoxin M₁ (III, R=OH),^{3,4} which is one of a



I

II

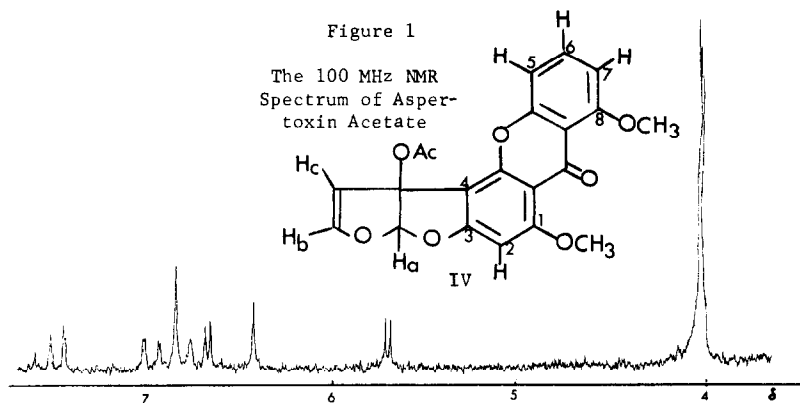
III

number of aflatoxins produced by the mold A. flavus. The obvious structural resemblance of aflatoxin B₁ (III, R=H)⁵ to a metabolite of the mold Aspergillus versicolor, sterigmatocystin (II, R=H),⁶ and the fact that the aflatoxins and sterigmatocystin are derived from the same genus, led to the early postulation that sterigmatocystin, or a sterigmatocystin precursor, may be a biosynthetic precursor of the aflatoxins.⁷ The very recent discovery that a simple sterigmatocystin derivative, O-methylsterigmatocystin (II, R=CH₃),² is elaborated by an

aflatoxin-producing strain of *A. flavus*, and the present report that aspertoxin is the second sterigmatocystin derivative to be isolated from *A. flavus* cultures, furnish support for the above hypothesis.

Aspertoxin is a colorless, crystalline material which undergoes decomposition, without melting, in the range 240-280° and affords a complex mixture (TLC) of highly colored products. The molecular formula of aspertoxin was established by high resolution mass spectrometry; the compound exhibited a molecular ion peak at $m/e = 354.074200$, and the mass calculated for $C_{19}H_{14}O_7$ is 354.073949. Elemental analysis indicates that aspertoxin crystallizes as the hydrate (Calcd. for $C_{19}H_{14}O_7 \cdot H_2O$: C, 61.29; H, 4.33; Found: C, 61.20; H, 4.45). Treatment of aspertoxin with acetic anhydride in pyridine at room temperature yielded aspertoxin acetate (IV), $C_{21}H_{16}O_8$, m.p. 119-121°. The mass spectrum of IV showed the molecular ion peak at $m/e = 396$.

The NMR spectrum of aspertoxin acetate (IV) is presented as Figure 1. The signal due to



the acetyl protons is not shown in Figure 1, and appears as a singlet at $\delta 2.17(3H)$. Signals due to the methoxyl hydrogens are prominent at $\delta 4.07$ and $\delta 4.10(6H)$. The argument for the attachment of the four carbon side chain at C_4 of the xanthone ring rather than at C_2 is based on the chemical shift observed for the isolated aromatic proton at C_2 (see IV, above). This proton gives a singlet at $\delta 6.41(1H)$, and the position of this signal agrees closely with the chemical shifts observed for the corresponding aromatic protons in *O*-methylsterigmatocystin (II, $R=CH_3$, $\delta 6.38$)² and sterigmatocystin (II, $R=H$, $\delta 6.45$).⁶

Signals for the three protons in aspertoxin acetate (IV) at $\delta 5.66$ (doublet, $J \approx 2$ Hz), $\delta 6.64$ (doublet, $J \approx 2$ Hz), and $\delta 6.83$ (singlet) may be attributed to protons H_c , H_b and H_a ,

respectively. The coupling between protons H_b and H_c was demonstrated by a double irradiation experiment. The position and coupling of the two vinyl protons (H_b and H_c), and the position of the acetal proton (H_a) signal are similar to those observed for the corresponding protons in aflatoxin M_1 (III, R=OH).⁴ The two vinyl protons in aflatoxin M_1 appear as doublets ($J \approx 2$ Hz) at $\approx \delta 5.8$ and $\approx \delta 6.9$. The single acetal proton in aflatoxin M_1 gives a singlet at $\approx \delta 6.6$.

The remaining three aromatic protons H_5 , H_6 and H_7 give rise to an ABC spectrum which can be analyzed roughly by the ABX approximation. The X portion consists of a triplet* at $\delta 7.52$ with a spacing of about 8 Hz. The AB portion cannot be seen in its entirety, since the acetal proton (H_a) peak is superimposed on it. The discernible absorptions are three doublets with a spacing of about 1 Hz, whereas the fourth such doublet seems to be merged with the acetal proton peak. With this assumption, the analysis for the ABX pattern yields the following approximate parameters: $J_{AB} = 1$ Hz; $J_{AX} = J_{BX} = 8$ Hz; $\nu_A = 698$ Hz and $\nu_B = 678$ Hz (measured downfield from TMS). The appearance of the X portion as a triplet is typical for $J_{AX} = J_{BX}$. That $\nu_A - \nu_B \neq 0$ was borne out further by the results of strong irradiation at the frequency of the center of the $\delta 7.52$ triplet: the AB portion changed into two signals at $\delta 6.98$ and $\delta 6.78$, their position corresponding to ν_A and ν_B as found above. The assignment of A, B and X to H_5 , H_6 and H_7 is then as follows: (A,B) = (H_5, H_7), and X = H_6 . The values for $J_{AX} = J_{BX}$ (8 Hz) and J_{AB} (1 Hz) fall in the ranges for J_{ortho} and J_{meta} in benzenoid systems. It should be mentioned also that the present ABX pattern bears a striking resemblance to that of the corresponding protons in O-methylsterigmatocystin (II, R= CH_3)² which consists of a triplet at $\delta 7.48$ (1 H) with spacings of 8.1 Hz and a complex band centered at $\delta 6.82$ (2H).

The U.V. spectrum of aspertoxin has λ_{max}^{MeOH} at 310 $m\mu$ and 241 $m\mu$ ($\log \epsilon$ 4.08 and 4.53, respectively). The U.V. spectrum of O-methylsterigmatocystin has λ_{max}^{MeOH} at 310 $m\mu$ and 236 $m\mu$ ($\log \epsilon$ 4.224 and 4.614, respectively).² The U.V. spectrum of aspertoxin is unaffected by the addition of acid or base.

The I.R. spectrum of aspertoxin (KBr disc) shows strong absorptions at 3354 cm^{-1} (OH), 1656 (γ -pyrone carbonyl), 1630, 1587, 1575, (phenyl), 1460, 1269, 1133 (C-O-C), 894, and 811 cm^{-1} (isolated aromatic H). The I.R. spectrum of aspertoxin acetate (in $CHBr_3$) is compared with the reported I.R. spectrum of O-methylsterigmatocystin (in $CHBr_3$)⁸ in Table I.

* The tall peak at $\delta 7.45$ next to the highest-field component of the triplet is due to $CHCl_3$.

The shapes and relative intensities of the major peaks in the spectrum of O-methylsterigmatocystin² are remarkably similar to those of aspertoxin and aspertoxin acetate.

TABLE I
COMPARISON OF I.R. SPECTRA OF O-METHYLSTERIGMATOCYSTIN⁸ AND ASPERTO-
XIN ACETATE. ν_{MAX} IN CM^{-1} (S = STRONG, M = MEDIUM, W = WEAK).

ASPERTOIN ACETATE (IV)			O-METHYLSTERIGMATOCYSTIN (II, R=CH ₃)		
1739 s	1414 m	1083 m	---	1418 m	1075 m
1655 s	1380 w	---	1662 s	1382 w	1040 w
1640 s	1341 m	1011 m	1643 s	1347 m	1016 m
1617 (sh)	1262 s	965 m	*	1267 s	970 m
1592 s	---	890 w	1603 s	1250 m	890 w
1469 s	1230 s	850 w	1473 s	1229 m	840 w
1430 m	---	811 m	1438 m	1204 m	811 m

*Ref. 9 does not list an absorption near 1617 cm^{-1} ; however, Ref. 2, which presents a photograph of the I. R. spectrum of O-methylsterigmatocystin, shows that the compound does indeed have a shoulder absorption near 1620 cm^{-1} .

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